## **EDITORIAL**

## The Road Traveled: Genomics and Biomarkers in Antineutrophil Cytoplasmic Antibody–Associated Vasculitis

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The search for biomarkers of disease activity and response to therapy is a continuing quest, and the increasing sophistication of genomic data, including transcriptomics, and of capacities for high-dimensional data analytics point to encouraging opportunities on the horizon for many diseases. In this issue of *Arthritis & Rheumatology*, Grayson et al (1) leveraged biologic specimens gathered during the Rituximab in ANCA-Associated Vasculitis (RAVE) trial and applied RNA sequencing to samples of whole blood and separated leukocytes to search for gene expression patterns that might serve as biomarkers that are potentially involved in pathophysiology and response to therapy.

Similar to several other autoimmune diseases with prominent neutrophil-related gene expression signatures (2–5), AAV patients in the RAVE trial had a discernable neutrophil-related profile in the transcriptomes of their circulating leukocytes. A lower intensity of the granulocyte multigene composite score, which was created for this study, was related to a higher likelihood of meeting the primary end point in the trial. Taken in the context of other clinical studies demonstrating neutrophil-related gene expression signatures in AAV, the data reported by Grayson et al further support a potential role of neutrophils and, likely, low-density granulocytes (LDGs) in the pathophysiology of AAV and a role of neutrophil markers in informing therapeutic strategies.

As with all studies at the leading edge of clinical research, this study prompts consideration of a series of opportunities for methodologic refinement and for replication in independent data sets. The authors acknowledge that the use of whole blood, even when depleted of globin

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Submitted for publication March 25, 2015; accepted in revised form April 7, 2015.

transcripts, limits the ability to detect less-common transcripts and may introduce confounding due to variations across participants in the proportion and number of different circulating cell types, each with characteristic gene expression signatures. Nonetheless, it would be interesting to use unsupervised hierarchical clustering as in this study to generate additional granulocyte multigene composite scores derived from other AAV data sets and then compare the performance of such scoring systems in relation to clinical outcomes for the RAVE participants. Agreement on a composite score derived from multiple different training data sets and applied successfully to several different independent sets of outcome data, as is being pursued in other diseases (6), would enhance the utility of a transcriptomic approach to practicable biomarkers. Furthermore, in future longitudinal studies, the performance of granulocyte-related gene expression profiles as predictors of therapeutic responsiveness could be compared with absolute neutrophil counts and flow cytometric measurements of circulating low-density granulocytes (frequency and absolute numbers) as defined by size, granularity, and CD15 and CD16 expression (7). Such an approach would provide further insights into both pathophysiology and real-time clinical feasibility.

An AAV cohort study of neutrophil properties, including membrane proteins and apoptosis, is currently under way in Europe at the Hôpital Cochin (Assistance Publique-Hôpitaux de Paris; ClinicalTrials.gov identifier: NCT01862068) and might provide additional such data. Similar future studies might also include other variables, such as ANCA titer and immunoglobulin class (8), as well as genetic variants that have been implicated in both the pathogenesis and clinical course of AAV (8–14). Since such high-dimensionality data will require large numbers of participants for sufficient statistical power, a cooperative effort across patients, health care professionals, academic institutions, and sponsoring agencies is likely to be necessary. Nonetheless, the opportunity for more individualized evaluation of our patients and more precise use of our therapeutic armamentarium—the EDITORIAL 1701

right therapy at the right time—is both exciting and compelling in the advancement of clinical care.

Insights into pathogenesis that prompted this study are provocative. The ability of ANCA to activate neutrophils with surface proteinase 3 (PR3) for further degranulation and formation of neutrophil extracellular traps (NETs) (8,9,15) may synergize with the spontaneous ability of low-density granulocytes (LDGs), which have now been formally demonstrated in AAV patients to form NETs. The process of perivascular NETosis may enhance vascular injury characteristic of AAV (5,7). While the genesis of circulating LDGs may involve premature release of an immature granulocyte subset from the marrow (7), it may also be that some LDGs represent stimulated, partially degranulated, more-mature neutrophils. Development of a neutrophil-oriented therapeutic strategy might depend on the genesis of, and the role played by, LDGs in AAV, and investigations could approach the hypothesis that LDGs represent stimulated, partially degranulated, more-mature neutrophils by looking for surface PR3 and CD177 on LDGs from AAV patients, as these are the surface constituents that facilitate ANCA-mediated stimulation of neutrophils (16,17).

It might also be insightful to compare the gene expression profiles of LDGs obtained from patients with different diseases with the question of whether differences in these profiles could provide important clues to pathogenesis. Interestingly, 41 of the 281 genes that defined an LDG signature in a lupus cohort (5) overlapped with genes differentially expressed in the RAVE nonresponders. While the 41 overlapping genes support some commonality in the biology of LDGs in different diseases, perhaps the 240 nonoverlapping genes have another story to tell. They may provide insight into the development of LDGs, the heterogeneity of LDGs, and perhaps the different stimuli encountered in the *milieu interieur* of each disease.

The role of neutrophils—and potentially of LDGs—in the pathogenesis of AAV raises several therapeutic challenges. Effective targeting of neutrophils in animal models, especially with depletion strategies, has been in large part problematic because of an impaired host defense when a critical "first responder" cell type is unavailable. The increased infection risk in leukocyte adhesion deficiency syndromes in humans (18) suggests that targeting myeloid cell margination and adhesion, at least chronically, may also have problematic safety profiles. However, an alternative might be to consider compounds such as glyburide, a K<sub>ATP</sub>-channel blocker and broad-spectrum ATP-binding cassette transporter inhibitor used to treat type 2 diabetes mellitus. Some evidence indicates that glyburide is anti-

inflammatory and may down-regulate the expression of neutrophil-related genes related to AAV pathogenesis, including CD177, CD89, and CD64 (see Supplementary Table 3A in ref. 19).

Whether such a repurposing strategy with glyburide or similar compounds would be effective with LDGs, as opposed to mature neutrophils, and effective in reducing neutrophil-mediated tissue injury in AAV are questions for future investigation. Nonetheless, the clear presence of a neutrophil gene expression signature, the role of ANCA and ANCA target on the neutrophil surface, and the efficacy of rituximab suggest a complexity in AAV pathogenesis that will likely require a multifaceted approach.

The work by the RAVE-Immune Tolerance Network (RAVE-ITN) Research Group encourages us to consider genomics, in addition to genetics and the many parameters of an activated immune system, in both the pathogenesis and the management of AAV. Increasingly, genetics is assuming a role in precision medicine, and we can anticipate that, as functional genomics provides additional insight into pathophysiology, it will also contribute to the portfolio of biomarkers that inform our therapeutic decisions. There is much work to be done, but the road to be traveled with our patients will take us to exciting insights into the mechanisms and better treatments for the diseases that challenge us each day.

## **AUTHOR CONTRIBUTIONS**

Dr. Kimberly drafted the article, revised it critically for important intellectual content, and approved the final version to be published.

## REFERENCES

- Grayson PC, Carmona-Rivera C, Xu L, Lim N, Gao Z, Asare AL, et al, for the Rituximab in ANCA-Associated Vasculitis– Immune Tolerance Network Research Group. Neutrophilrelated gene expression and low-density granulocytes associated with disease activity and response to treatment in antineutrophil cytoplasmic antibody-associated vasculitis. Arthritis Rheumatol 2015;67:1922-32.
- Cheadle C, Berger AE, Andrade F, James R, Johnson K, Watkins T, et al. Transcription of proteinase 3 and related myelopoiesis genes in peripheral blood mononuclear cells of patients with active Wegener's granulomatosis. Arthritis Rheum 2010;62:1744–54.
- 3. Lyons PA, McKinney EF, Rayner TF, Hatton A, Woffendin HB, Koukoulaki M, et al. Novel expression signatures identified by transcriptional analysis of separated leucocyte subsets in systemic lupus erythematosus and vasculitis. Ann Rheum Dis 2010;69:1208–13.
- Bennett L, Palucka AK, Arce E, Cantrell V, Borvak J, Banchereau J, et al. Interferon and granulopoiesis signatures in systemic lupus erythematosus blood. J Exp Med 2003;197:711–23.
- Villanueva E, Yalavarthi S, Berthier CC, Hodgin JB, Khandpur R, Lin AM, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. J Immunol 2011;187:538–52.
- Chiche L, Jourde-Chiche N, Whalen E, Presnell S, Gersuk V, Dang K, et al. Modular transcriptional repertoire analyses of

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- adults with systemic lupus erythematosus reveal distinct type I and type II interferon signatures. Arthritis Rheumatol 2014;66: 1583–95.
- Carmona-Rivera C, Kaplan MJ. Low-density granulocytes: a distinct class of neutrophils in systemic autoimmunity. Semin Immunopathol 2013;35:455–63.
- Kelley JM, Monach PA, Ji C, Zhou Y, Wu J, Tanaka S, et al. IgA and IgG antineutrophil cytoplasmic antibody engagement of Fc receptor genetic variants influences granulomatosis with polyangiitis. Proc Natl Acad Sci U S A 2011;108:20736–41.
- Porges AJ, Redecha PB, Kimberly WT, Csernok E, Gross WL, Kimberly RP. Anti-neutrophil cytoplasmic antibodies engage and activate human neutrophils via FcγRIIa. J Immunol 1994;153: 1271–80.
- Kocher M, Edberg JC, Fleit HB, Kimberly RP. Antineutrophil cytoplasmic antibodies preferentially engage FcγRIIIb on human neutrophils. J Immunol 1998;161:6909–14.
- Mahr AD, Edberg JC, Stone JH, Hoffman GS, St.Clair EW, Specks U, et al, for the Wegener's Granulomatosis Genetic Repository Research Group. Alpha<sub>1</sub>-antitrypsin deficiency– related alleles Z and S and the risk of Wegener's granulomatosis. Arthritis Rheum 2010;62:3760–7.
- Lyons PA, Rayner TF, Trivedi S, Holle JU, Watts RA, Jayne DR, et al. Genetically distinct subsets within ANCA-associated vasculitis. N Engl J Med 2012;367:214–23.

- Carney EF. Vasculitis syndromes: AAV encompasses two major genetically distinct conditions with different autoantibody specificities. Nat Rev Rheumatol 2012;8:502.
- Alberici F, Martorana D, Vaglio A. Genetic aspects of antineutrophil cytoplasmic antibody-associated vasculitis. Nephrol Dial Transplant 2015;30 Suppl 1:i37–45.
- Kessenbrock K, Krumbholz M, Schonermarck U, Back W, Gross WL, Werb Z, et al. Netting neutrophils in autoimmune smallvessel vasculitis. Nat Med 2009;15:623–5.
- 16. Hu N, Westra J, Huitema MG, Bijl M, Brouwer E, Stegeman CA, et al. Coexpression of CD177 and membrane proteinase 3 on neutrophils in antineutrophil cytoplasmic autoantibody-associated systemic vasculitis: anti-proteinase 3-mediated neutrophil activation is independent of the role of CD177-expressing neutrophils. Arthritis Rheum 2009;60:1548-57.
- Jerke U, Rolle S, Dittmar G, Bayat B, Santoso S, Sporbert A, et al. Complement receptor Mac-1 is an adaptor for NB1 (CD177)-mediated PR3-ANCA neutrophil activation. J Biol Chem 2011;286:7070–81.
- 18. Van de Vijver E, van den Berg TK, Kuijpers TW. Leukocyte adhesion deficiencies. Hematol Oncol Clin North Am 2013;27:101–16.
- Koh GC, Maude RR, Schreiber MF, Limmathurotsakul D, Wiersinga WJ, Wuthiekanun V, et al. Glyburide is antiinflammatory and associated with reduced mortality in melioidosis. Clin Infect Dis 2011;52:717–25.